

Genetic and morphological divergence between *Littorina fabalis* ecotypes in Northern Europe

Juan Galindo^{1,2*}  | João Carvalho^{3*}  | Graciela Sotelo^{4,5} | Mårten Duvetorp⁶ | Diana Costa^{4,7}  | Petri Kempainen⁸ | Marina Panova⁶ | Antigoni Kaliontzopoulou⁴  | Kerstin Johannesson⁶  | Rui Faria^{4,5,7} 

¹Department of Biochemistry, Genetics and Immunology, Universidade de Vigo, Vigo, Spain

²Centro de Investigación Mariña (CIM-UVIGO), Vigo, Spain

³e3c – Centre for Ecology, Evolution and Environmental Changes, Faculdade de Ciências, Universidade de Lisboa, Campo Grande, Portugal

⁴CIBIO, Centro de Investigação em Biodiversidade e Recursos Genéticos, InBIO, Laboratório Associado, Universidade do Porto, Vairão, Portugal

⁵Department of Animal and Plant Sciences, University of Sheffield, Sheffield, UK

⁶Department of Marine Sciences –Tjärnö, University of Gothenburg, Strömstad, Sweden

⁷CIIMAR, Interdisciplinary Centre of Marine and Environmental Research, University of Porto, Porto, Portugal

⁸Ecological Genetics Research Unit, Research Programme in Organismal and Evolutionary Biology, Faculty of Biological and Environmental Sciences, University of Helsinki, Helsinki, Finland

Correspondence

Rui Faria, CIBIO, Centro de Investigação em Biodiversidade e Recursos Genéticos, Universidade do Porto, 4485-661 Vairão, Portugal.

Email: r.macieiradefaria@sheffield.ac.uk

Juan Galindo, Department of Biochemistry, Genetics and Immunology, University of Vigo, Vigo, Spain.

Email: galindo@uvigo.es

Funding information

Volkswagen Stiftung, Grant/Award Number: 50500776; European Union, Grant/Award Number: 706376; European Marine Biological Resource Centre, Grant/Award Number: 227799; Fundação para a Ciência e a Tecnologia, Grant/Award Number: PTDC/BIA-EVF/113805/2009, IF/00641/2014/CP1256/CT0008, PTDC/BIA-EVL/30628/2017, PD/BD/128350/2017 and IF/00641/2014/CP1256/CT0008; European Commission, Grant/Award Number: FCOMP-01-0124-FEDER-014272 and POCI-01-0145-FEDER-030628; Ministerio de Ciencia, Innovación y Universidades, Spain, Grant/Award Number: RTI2018-101274-J-I00

Abstract

Low dispersal marine intertidal species facing strong divergent selective pressures associated with steep environmental gradients have a great potential to inform us about local adaptation and reproductive isolation. Among these, gastropods of the genus *Littorina* offer a unique system to study parallel phenotypic divergence resulting from adaptation to different habitats related with wave exposure. In this study, we focused on two *Littorina fabalis* ecotypes from Northern European shores and compared patterns of habitat-related phenotypic and genetic divergence across three different geographic levels (local, regional and global). Geometric morphometric analyses revealed that individuals from habitats moderately exposed to waves usually present a larger shell size with a wider aperture than those from sheltered habitats. The phenotypic clustering of *L. fabalis* by habitat across most locations (mainly in terms of shell size) support an important role of ecology in morphological divergence. A genome scan based on amplified fragment length polymorphisms (AFLPs) revealed a heterogeneous pattern of differentiation across the genome between populations from the two different habitats, suggesting ecotype divergence in the presence of gene flow. The contrasting patterns of genetic structure between nonoutlier and outlier loci, and the decreased sharing of outlier loci with geographic distance among locations are compatible with parallel evolution of phenotypic divergence, with an important contribution of gene flow and/or ancestral variation. In the

*Juan Galindo and João Carvalho have equal contribution.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. *Journal of Evolutionary Biology* published by John Wiley & Sons Ltd on behalf of European Society for Evolutionary Biology

future, model-based inference studies based on sequence data across the entire genome will help unravelling these evolutionary hypotheses, improving our knowledge about adaptation and its influence on diversification within the marine realm.

KEYWORDS

adaptation, AFLPs, divergent natural selection, ecological speciation, flat periwinkles, gene flow, geometrics–morphometrics, parallel evolution, shell morphology

1 | INTRODUCTION

The marine rocky intertidal represents one of the most abrupt environmental gradients on Earth (Little & Kitching, 1996; Raffaelli & Hawkins, 1996; Tomanek & Helmuth, 2002). Different environmental conditions across the tidal range result in patterns of vertical and horizontal (along the shore) zonation both in terms of species diversity and in terms of intraspecific phenotypic variation (Connell, 1972; Little & Kitching, 1996; Raffaelli & Hawkins, 1996; Southward, 1957). Wave action is one of the most important physical selective agents across intertidal environments worldwide, shaping both axes of zonation (Le Pennec et al., 2017). Habitats exposed to, or sheltered from waves differ consistently in their biodiversity (Denny & Wetthey, 2001; Helmuth & Denny, 2003). Biotic factors are also known to affect the intertidal community (e.g. presence of predators; Paine et al., 1994), which adds to the abiotic selective pressures (Seeley, 1986). Altogether, these environmental gradients make the intertidal a natural laboratory to study local adaptation and ecological speciation.

Taxa with low dispersal, where divergent selection can be strong enough to counteract gene flow among populations and promote divergence with gene flow, are particularly well suited for studies about local adaptation and ecological speciation (Sanford & Kelly, 2011; Smadja & Butlin, 2011). Among these, species with distinctive phenotypes associated with different microhabitats (i.e. ecotypes) in the intertidal can provide important information about how natural selection influences biological diversification (e.g. Coyer et al., 2011; Kess et al., 2018; Sanford & Kelly, 2011; Wilding et al., 2001). Instances of parallel evolution of ecotypes across similar environmental gradients in multiple locations across a species' distribution, that is parallel evolution, are viewed as support for a role of natural selection in driving divergence. This is because it is unlikely that ecotypes have repeatedly evolved in the same phenotypic direction if only purely stochastic processes were involved (Johannesson, 2001; Nosil, 2012; Schluter, 2000).

The characterization of the genetic variation underlying parallel evolution allows to distinguish if repeated events of phenotypic divergence tend to involve the same *de novo* mutations, different *de novo* mutations in the same or different genomic regions, ancestrally shared standing polymorphisms, the same alleles due to migration between populations or a combination of all the above (Elmer & Meyer, 2011; Faria et al., 2014; Johannesson et al., 2010;

Nosil, 2012). Thus, the study of parallel phenotypic divergence across intertidal microhabitats can help us understand if adaptation usually involves the same genetic paths, as well as the relative contributions of ancestral polymorphism and/or gene flow.

Parallel ecotypic divergence in the intertidal zone has been particularly well documented in the rough periwinkle *Littorina saxatilis* (Gastropoda; Galindo & Grahame, 2014; Reid, 1996; Rolán-Alvarez et al., 2015). Two main ecotypes have been described: a large, thick-shelled ecotype inhabiting sheltered microhabitats that faces intense crab predation (Crab ecotype); and a small, thin-shelled one facing heavy surf in exposed microhabitats (Wave ecotype) (reviewed in Johannesson et al., 2010). These ecotypes, found only tens of metres apart in Spain, the United Kingdom (UK) and Sweden, have diverged in parallel within each of these countries (Butlin et al., 2014). Amplified fragment length polymorphisms (AFLPs) allowed the initial identification of loci under divergent selection (hereafter “outliers”) between ecotypes and provided the first insights on the proportion of outliers shared at different geographic scales, within and among countries (Galindo et al., 2009, 2013; Grahame et al., 2006; Hollander et al., 2015; Wilding et al., 2001). Benefiting from the assembly of a reference genome and the construction of a genetic map for *L. saxatilis*, the heterogeneous genomic differentiation between ecotypes was recently confirmed with the identification of some genomic regions enriched for the presence of outliers, which tend to coincide with polymorphic inversions (Faria et al., 2019; Westram et al., 2018). Moreover, whole-genome sequencing of pools of individuals from multiple populations of both ecotypes across the species' distribution range revealed that outlier sharing tends to be high even among distant populations, although it decreases with the geographic distance between populations (Morales et al., 2019).

A closely related species for which phenotypic variation is also found associated with an environmental cline in wave exposure is *Littorina fabalis* (Kemppainen et al., 2005; Reimchen, 1981; Tatarenkov & Johannesson, 1994, 1998, 1999). Individuals with large and thick shells (hereafter “large ecotype”) are commonly found in moderately exposed shores, whereas individuals with smaller and thinner shells (hereafter “dwarf ecotype”) predominate in sheltered habitats. Contrary to *L. saxatilis* that live on rocks, these two *L. fabalis* ecotypes dwell on brown macroalgae (*Fucus* spp. and *Ascophyllum* spp.), grazing on the epiphytes that grow on the algae fronds (Williams, 1990). The fronds are also thought to provide refuge against one of their main predators, the green crabs

(*Carcinus maenas*), that are found in both microhabitats (Kempainen et al., 2005).

Habitat-related variation in one allozyme locus (arginine kinase, *Ark*) was initially found in Swedish populations of *L. fabalis*, suggesting that this locus was under the influence of natural selection related to wave exposure and/or other associated factors (Tatarenkov & Johannesson, 1994). The differences in *Ark* allele frequencies between sheltered and moderately exposed habitats were also associated with variation at a random amplification of polymorphic DNA (RAPD) locus and with the size differences described above. This is true even for sites with intermediate exposure, suggesting some reproductive isolation between the ecotypes despite gene flow (Johannesson & Mikhailova, 2004; Tatarenkov & Johannesson, 1998).

Similar habitat-related phenotypic divergence has also been observed at least in the UK (Wales), France and Norway, where contrary to Sweden, *L. fabalis* also has to face high tidal amplitudes (Kempainen et al., 2009, 2011; Reimchen, 1981; Tatarenkov & Johannesson, 1999). Even if a northern refugium could have existed during the last glacial maximum (LGM) for *L. fabalis* (Sotelo et al., 2020), most of these shores were likely colonized after the LGM (Charbit et al., 2007). This, together with the absence of significant genetic differentiation between the ecotypes across this region using neutral markers (Sotelo et al., 2020), suggests a relatively recent (after the LGM) local establishment of habitat-related phenotypic divergence.

Contrary to neutral markers, *Ark* intron sequencing revealed highly significant divergence between individuals from sheltered and moderately exposed habitats (Kempainen et al., 2011). Results show that one haplotype was almost fixed and shared across sheltered habitats of these different countries whereas wave-exposed habitats maintained similarly higher variation. This increases the possibility of “evolution in concert”, where locally adapted alleles could arise once and subsequently spread to geographically distant populations inhabiting the same habitat by means of ecotype-specific selective sweeps (Johannesson et al., 2010; Kempainen et al., 2011; Schluter & Conte, 2009). However, except for *Ark* and the putatively linked RAPD locus, whether the same genetic variation is involved in the evolution of these ecotypes is currently unknown. Thus, studies that integrate both morphological data and a high number of nuclear markers from different locations are needed to evaluate the parallelism of phenotypic divergence in *L. fabalis* across its geographic range, as well as the genetic variation and processes involved.

In this study, we used shell geometric morphometrics and AFLPs to perform the first characterization of *L. fabalis* ecotypes across multiple Northern European populations (Norway, Sweden and the UK) with four main goals: (a) to understand if shell shape and size divergence evolved in the same direction among locations within the same country and across countries; (b) to identify loci involved in the differentiation between sheltered and exposed sites (i.e. outlier loci); (c) to quantify the degree of outlier sharing at two different geographic scales: within and among countries; and (d) to contrast

population structure and relatedness using outlier (putatively adaptive) versus nonoutlier loci (putatively neutral) in order to gain insights about the evolutionary history of phenotypic divergence. Although not fully conclusive, our results are compatible with phenotypic parallel divergence within *L. fabalis* associated with habitat and reveal a pattern of outlier sharing that suggests a relevant role of gene flow and/or retention of ancestral polymorphism in the evolution of ecotypes.

2 | MATERIAL AND METHODS

2.1 | Sampling

Littorina fabalis individuals from both sheltered and moderately exposed habitats were randomly collected with respect to their shell morphology between August and October 2012, the sample size in each population varied between 21 and 72 (average $N = 39$) (Table S1). A nested sampling design was implemented: samples were collected from each habitat within each location with replicates at two geographical scales: regional (two to three locations within each country, <50 km) and global (different countries, >1,000 km) (Figure 1, Figure S1, Table S1). This allows the investigation of parallel evolution at these two contrasting scales.

Within each location, sites representing each type of habitat were preselected based on information from previous studies (Kempainen et al., 2009; Tatarenkov & Johannesson, 1994), or on their orientation and topography as retrieved from the Google Earth Engine (Gorelick et al., 2017). This preclassification was further confirmed in situ, where sheltered habitats were distinguished from moderately exposed ones by the high abundance of *Ascophyllum* spp., a macroalga that is commonly used as an indicator of sheltered shores (Tatarenkov & Johannesson, 1998). However, two putatively exposed sites did not adjust to these patterns: those in Anglesey South (UK) and in Ursholmen (Sweden) (Table S1). In Anglesey South, *Ascophyllum* spp. was only abundant in the upper part of the shore but individuals were collected from the lower part, where wave action is likely rather high. In Ursholmen, the moderately exposed site was chosen based on its orientation towards relatively open sea but *Ascophyllum* spp. was highly abundant there. The site was still included in the study but re-classified as sheltered (Table S1). Since we were not able to sample a proper moderately exposed site in Ursholmen, this location was excluded from statistical tests on shell morphology. It is also important to emphasize that the distance between habitats within each location differs among countries. In the Scandinavian locations, snails are continuously distributed along the shore from sheltered to moderately exposed sites (i.e. parapatric), which are <500 m apart. In the UK, the two habitats within each sampled location are geographically isolated from each other, from 400 m to 8 km apart (Figure 1, Figure S1).

After collection, individuals were brought alive to the laboratory where they were frozen at -20°C and then stored in 95% ethanol.

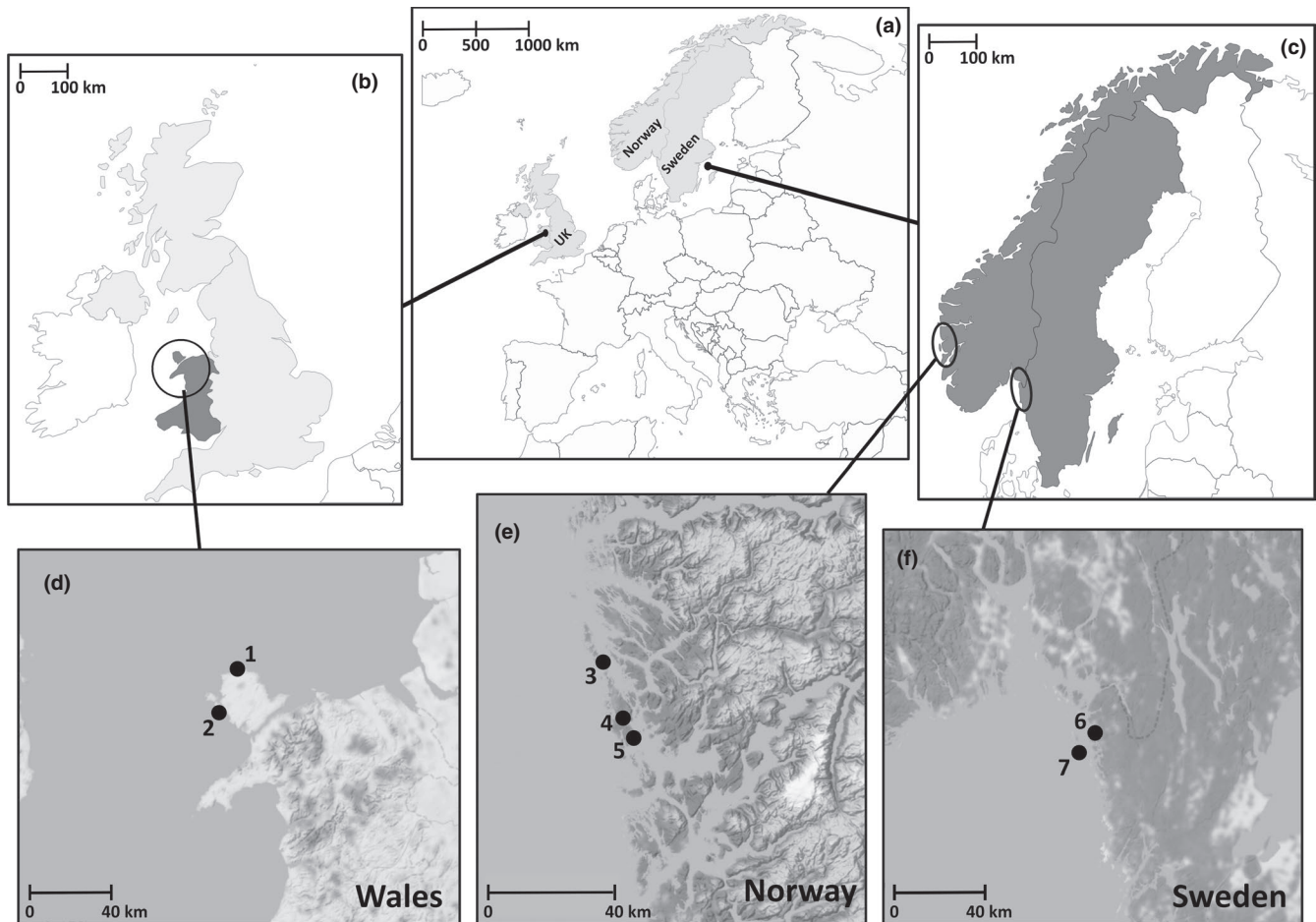


FIGURE 1 Sampling locations. (a) Map of Europe highlighting the three countries where populations of *Littorina fabalis* were sampled: (b) Wales, United Kingdom (UK), (c) Norway and Sweden. Zoom in showing the sampling locations in Wales (d), Norway (e) and Sweden (f) 1-Anglesey North and 2-Anglesey South, in Wales; 3-Seløyna, 4-Syltøyna and 5-Hummelsund, in Norway; 6-Lökholmen and 7-Ursholmen, in Sweden.

2.2 | Sample processing and classification into species

Shells of adult individuals were photographed over graph paper (used for scale) in a standardized position following Carvajal-Rodríguez et al. (2005) and using a digital ICA video module fitted on a MZ12 dissection microscope (Leica) for subsequent geometric morphometric analyses. Shells were then broken, and individuals dissected and sexed under the same microscope. Since shell morphology is not completely diagnostic between the two closely related species of flat periwinkles (*L. fabalis* and *L. obtusata*), we could not guarantee that all collected individuals were indeed *L. fabalis* just based on their shells. Thus, we focused our analysis on males, which were classified into *L. fabalis* or *L. obtusata* based on the morphology of their genitalia, one of the most distinctive traits between the two species (Costa et al., 2020; Reid, 1996). In sites where the number of males was too low, females and immature individuals were also included. Nevertheless, all samples (including males) were later classified into species using the AFLP

genotypes. To do so, 17–19 individuals from *L. obtusata* populations within each country were deliberately included in the genetic analysis as references.

2.3 | Shell geometric morphometrics

Geometric morphometric (GM) analyses were carried out following the methodology previously developed for flat periwinkles by Costa et al. (2020). This consisted in digitizing a total of 28 landmarks for each shell of adult individuals (males and females classified as *L. fabalis*), including 4 fixed and 24 sliding semilandmarks, using TPSDIG v1.40 (Rohlf, 2006).

Superimposition of landmark coordinates was performed using generalized Procrustes analysis (GPA; Rohlf & Slice, 1990), retrieving shape variables (i.e. Procrustes residuals) and centroid size (CS), the latter used as an estimate of size for each individual. The final dataset consisted of 318 adult individuals (confirmed to be *L. fabalis* based on genitalia morphology and/or genetics), representing all seven locations.

A principal component analysis (PCA) of shape variables was conducted to assess overall patterns of variation. In addition, UPGMA dendrograms for size and shape were generated to evaluate clustering patterns between samples from each habitat and location, based on Euclidean distances (for size) and Procrustes distances (for shape) of population means. General linear models (GLMs) were then used to assess if shell shape and size (logCS) differed significantly between the two sampled habitats and to evaluate whether these differences varied among locations and countries. As such, GLMs included country, habitat and sampling location (nested within country) as main effects, as well as all interaction terms. Allometric effects were also investigated by performing a GLM on shape with logCS as a covariate, using the same factorial design. These analyses were carried out in the *geomorph* R package (Adams et al., 2019), using randomized residual permutation procedures (RRPP) of 1,000 permutations and Z-scores for significance assessment (Collyer & Adams, 2018, 2019). Deformation grids were used to visualize differences in shape between *L. fabalis* individuals from the two habitats within each location.

Finally, a discriminant function analysis (DFA) based on shape was implemented using the R package MASS v7.3.50 to infer the probability of morphological assignment of individuals into moderately exposed or sheltered habitats. The DFA was constructed based on the 287 individuals from all locations under study except for Ursholmen, using a leave-one-out cross-validation procedure. The resulting morphological posterior probability (PP) assignments were then compared with the genetic membership coefficients obtained in STRUCTURE (see below) for the 83 individuals for which both genetic and phenotypic data were available. All morphological data analyses were carried out in the R language for statistical computing (R Development Team, 2019).

2.4 | Genetic analyses

Genomic DNA was extracted using the CTAB–chloroform protocol described in Galindo et al. (2009). DNA quantity and purity were assessed with a BioDrop μ lite (BioDrop) spectrophotometer and adjusted to a final concentration of 20 ng/ μ l for each sample.

The AFLP analysis was performed using a modified version of the protocol developed for *L. saxatilis* by Galindo et al. (2009). Briefly, 100 ng of DNA was digested in a final volume of 12 μ l with 4U EcoRI (New England Biolabs, NEB) and 2U MseI (NEB) in 1X Buffer EcoRI (NEB) supplemented with 0.03 μ g of BSA. After a 3.5-hr incubation at 37°C, the ligation of Eco and Mse adaptors was done by adding 3 μ l of a solution containing 5 pmol of each adaptor and 0.25 U T4 DNA ligase (Roche) in 1X ligase buffer. The samples were incubated for 16 hr at 16°C. In digestion and subsequent steps, all samples were randomly distributed across 96-well plates, and 15% of replicate samples were included. The preselective PCR was performed in 10 μ l final volume containing 2 μ l of a 1:4 dilution of the ligation product, 0.3 mM of dNTP mix (Applied Biosystems), 2 mM of MgCl₂, 5 pmol of each preselective primer (Eco + A, Mse + C) and

0.3 U of Taq polymerase (Bioline) in 1X PCR Buffer. The first selective PCR was performed on 1 μ l of 1:4 dilution of the preselective PCR using the same reaction mixture but with the addition of 4 pmol Eco + ACT (FAM labelled), 2.5 pmol Eco + AAG (NED) and 5 pmol Mse + CAA primers. A second selective PCR was performed with Eco + ACT, Eco + AAG and Mse + CAC. PCR conditions, adaptor and primer sequences are described in Galindo et al. (2009). Selective PCR products were analysed on an ABI 3130 sequencer (Applied Biosystems) at CACTI (Centro de Apoyo Científico y Tecnológico a la Investigación, University of Vigo) sequencing facility along with GeneScan 500ROX (Applied Biosystems). Electropherograms were initially analysed with PeakScanner v.2.0 (Applied Biosystems). Loci were manually assigned by defining bins from the overlapping electropherograms of all the samples in RawGeno v.2.0 (Arrigo et al., 2012). The error rate (9% on average overall primer combinations) was estimated with the R package AFLPtools (<https://github.com/genevalab/AFLPTools>) that follows AFLPScore methodology (Whitlock et al., 2008). The final genotypes were obtained using this same package.

In order to identify *L. obtusata* individuals that could have been erroneously classified as *L. fabalis* based on shell appearance (prior GM analyses), AFLP-SURV v.1.0 (Vekemans et al., 2002) was used to calculate 1-relatedness coefficient (Lynch & Milligan, 1994) matrix between pairs of individuals, and then, a multidimensional scaling analysis (MDS) was implemented in R (R Core Team, 2018). The individuals that clustered with the *L. obtusata* individuals were removed from all morphological and genetic analyses, resulting in a final dataset formed by 503 *L. fabalis* individuals.

2.4.1 | Detection of outlier loci

Two different methodologies were used to identify outlier loci between *L. fabalis* exposed and sheltered habitats, BAYESCAN v.2.1 (Foll & Gaggiotti, 2008) and DFDIST (Beaumont & Nichols, 1996). In both cases, the analyses were carried out independently within each location. As previously mentioned, both Ursholmen sites were likely sheltered. Thus, we do not expect to detect outliers related to the level of wave-exposure or associated factors. Nevertheless, outliers between the two sites were still estimated as a control.

BAYESCAN calculates population-specific and locus-specific F_{ST} coefficients and then estimates the posterior probabilities of two alternative models (including or excluding the effect of selection) for each locus using a reversible-jump Markov chain Monte Carlo (MCMC) approach. Ten pilot runs (10,000 iterations) were performed to tune the model parameters, followed by 400,000 iterations (100,000 as burn-in, 20 as thinning interval and 20,000 as sample size). Loci were identified as outliers when the posterior probability was higher than 0.75 (equivalent to a Bayes factor of 3 or greater) (see Foll & Gaggiotti, 2008). DFDIST simulates loci (200,000) under a neutral model of two symmetrical islands with a mean F_{ST} adjusted to the empirical F_{ST} between the ecotypes calculated from the AFLP loci. Those loci with F_{ST} values significantly greater than

the simulated neutral distribution (F_{ST} conditional on heterozygosity) were considered as outliers using two different stringency criteria ($p > .95$ and $p > .99$). This analysis was conducted using the same parameters as Galindo et al. (2009).

Simulation studies that compare BAYESCAN to other methodologies for outlier detection, including DFDIST, have shown that BAYESCAN is more conservative concerning the number of outliers detected (Pérez-Figueroa et al., 2010). In this study, all BAYESCAN outliers were consistently included within the set of DFDIST outliers ($p > .99$). Because the goal of this study was to focus on the genetic structure of outlier versus nonoutlier loci (Galindo et al., 2009) rather than follow-up specific outlier loci (e.g. Wood et al., 2008), we carried out subsequent genetic analyses using the set of DFDIST outliers. Since the number of outliers (using the $p > .99$ cut-off) within some locations was relatively low (see results), substructure analyses were based on a more lenient criterion ($p > .95$). However, a more stringent criterion ($p > .99$) was used to compare the amount of shared outliers among locations (conservative). On the other hand, in order to be conservative, we considered nonoutlier loci only those with $p < .80$ in the DFDIST analysis.

2.4.2 | Genetics substructure analysis

Genetic substructure analyses were performed with these two different sets of loci, all outliers (i.e. putatively under divergent selection) and nonoutliers (i.e. putatively neutral). In both cases, they represent the combination of loci detected in each locality. AFLP-SURV v.1.0 (Vekemans et al., 2002) was used to calculate population pairwise genetic differentiation (F_{ST}) and Nei's genetic distance (10,000 bootstrap) using Zhivotovsky's (1999) Bayesian approach. Neighbour-joining (NJ) trees were constructed based on Nei's genetic distance using the NEIGHBOR routine implemented in the PHYLIP package (Felsenstein, 2013). The CONSENSE routine in PHYLIP was used to determine the bootstrap percentage supporting each branch of the tree. Trees were visualized using FigTree v.1.4.4 (<http://tree.bio.ed.ac.uk/software/figtree/>). A Bayesian clustering analysis was carried out in STRUCTURE v.2.3.4 (Pritchard et al., 2000) under three different hierarchical levels: (a) global, including all sites; (b) regional, including all sites within a country; and (c) local, including both the sheltered and exposed site within a location. All analyses were performed with the outlier and nonoutlier data sets, considering only as outliers those detected within the respective hierarchical levels (i.e. for the global analysis using all detected outliers in all pairwise comparisons, for the regional analysis only those detected in the pairwise comparisons within the corresponding region/country, whereas for the local analysis using only the outliers detected in that specific location). The consistency of individuals' assignment based on outliers across the different hierarchical levels was assessed for each location using correlation tests (Pearson correlation coefficient (r)- Sokal & Rohlf, 1995), implemented in the R package Stats v.3.6.1. For each K -value, we analysed five replicate runs of 500,000 iterations (100,000 as burn-in), assuming an admixture model, correlated

allele frequencies and without prior population information. All analyses were carried out from $K = 1$ up to $K =$ number of sites plus one, depending on the hierarchical level analysed. The method developed by Evanno et al. (2005) implemented in STRUCTURE HARVESTER was employed to determine the best K .

3 | RESULTS

3.1 | Characterization of shell morphology and phenotypic divergence

Adult individuals from each habitat differed significantly in size, with individuals from moderately exposed habitats being consistently larger than those from sheltered habitats (Figure 2a, Table S2). Accordingly, the UPGMA dendrogram based on centroid size (CS) revealed two main groups (Figure 2b): individuals from sheltered habitats from all locations on one hand, and all moderately exposed ones on the other except Syltøyna (Norway), which clustered within the sheltered group. Within each group, there was no particular geographic trend except for the clustering of all Norwegian sheltered habitats together. Other significant factors affecting size revealed by the general linear model (GLM) analysis included location and the interaction between habitat and location, suggesting that size differences between habitats varied across locations. However, size differences were not significant between countries (Table S2).

The same factors influencing size also influence shape variation, as reflected by significant effects of habitat, location and their interaction, but not of country (Table S2). When considering allometric effects, we found a significant influence of size on overall shape. In addition, size seems to affect shape differently across locations (significant interaction between both effects) (Table S2). Taking this effect of size into account, the same effects of habitat, location and their interaction remained significant, suggesting that although size accounts for some of the observed shape variation, it is not the only factor influencing it.

The distribution of individuals in the morphospace of the two principal components of shape variation, which explained a total of 69.4% of variation (PC1: 42.1%; PC2: 27.3%), revealed that average shapes of individuals from the moderately exposed habitats tend to cluster in the lower-left quadrant, whereas the average shape of individuals from the sheltered habitats is found in the upper-right part, with the exception of Anglesey South (Figure 3a). This agrees with the GLM results that show habitat as the factor with the strongest effect (based on Z-scores) on shape variation (as well as on size), followed by location (Table S2). The UPGMA dendrogram based on shape revealed two main groups, one consisting on the moderately exposed habitat from Scandinavian locations except Syltøyna (as for size); and the second comprising the sheltered habitats from Scandinavian locations and Syltøyna as well (Figure 3b). Contrary to what was observed in terms of size, although populations within the UK clustered by habitat rather than by location, they were nested within the clade composed mostly by sheltered sites from other

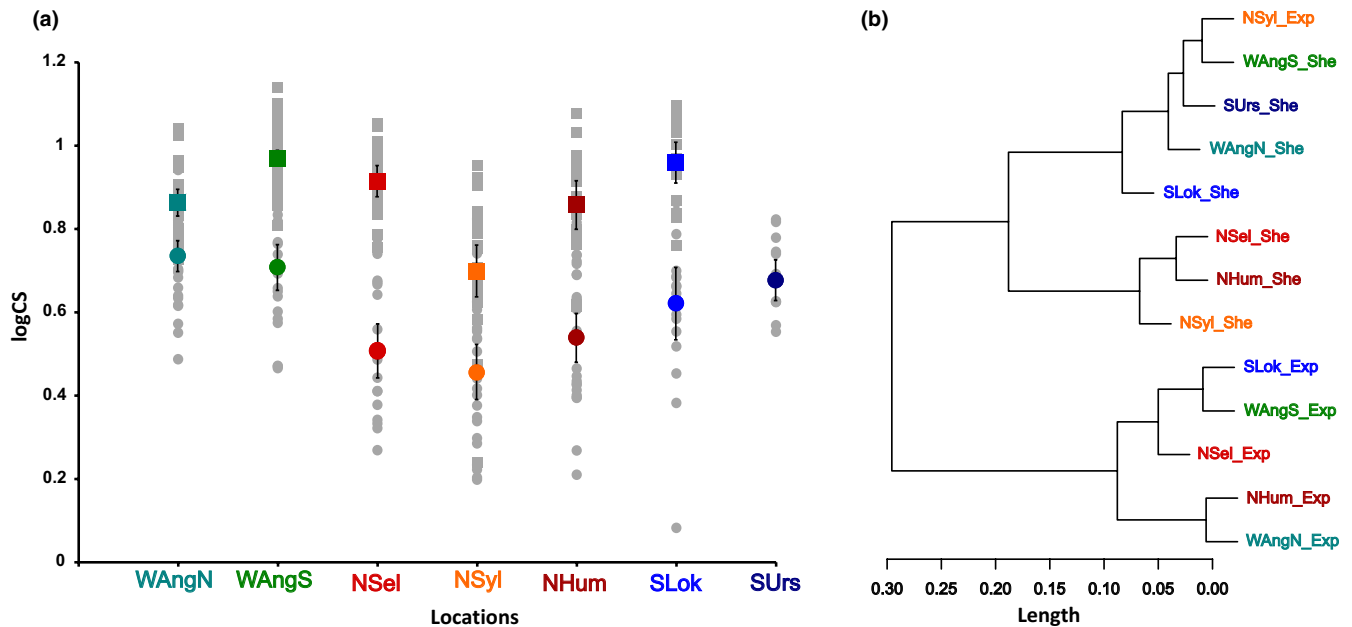


FIGURE 2 Results of the geometric morphometric (GM) analyses of shell size. (a) Mean size (logCS) of individuals for each habitat across locations. Mean values are indicated by coloured symbols (vertical bars denote 95% confidence intervals), whereas individuals are represented in grey; square and circle symbols represent the moderately exposed and sheltered habitats, respectively. (b) UPGMA dendrogram based on Euclidean distances of mean shell size. For Ursholmen, only samples of the sheltered habitat were included (see main text). Population codes are the same as in Table S1 and Figure S1

countries, suggesting also important geographic influence on shape, in accordance with GLM results (Table 2).

The inspection of deformation grids depicting the mean shape for individuals from each habitat (Figure 3c) shows that individuals inhabiting the moderately exposed habitat generally exhibit a larger width of the aperture (relative to overall shell size), whereas individuals of the sheltered habitat tend to exhibit a smaller aperture.

3.2 | Detection of outlier loci and comparison among locations

A total of 746 AFLP loci were analysed in 299 individuals (average sample size per population of 21) (Table S1). The lowest proportion of outliers was detected in the Swedish site Ursholmen (1.2%–5.3% for $p > .99$ and $p > .95$, respectively); and none of these outliers were shared with other locations (for $p > .99$), not even with the other Swedish location (Lökholmen, ~10 km apart) (Table 1). This suggests that indeed the populations from the two Ursholmen sites did not (or poorly) diverge according to the same axis of divergence (sheltered vs. moderately exposed) as in other locations, in agreement with the abundance of *Ascophyllum* spp. in both sampled habitats.

Excluding Ursholmen, a total of 1.6%–5.6% of outliers ($p > .99$) and 6.3%–9.5% of outliers ($p > .95$) were detected across all locations using DFDIST (Table 1, Figure S2). Using the most conservative threshold ($p > .99$), the number of outliers per location ranged from 6 in Lökholmen (Sweden) to 29 in Hummelsund (Norway) with the F_{ST} between habitats for these loci ranging from 0.0895 to 0.2922, respectively. This contrasts with lack of differentiation for nonoutlier

loci (Table 1). In total, only three outliers (3.8%) are shared among all countries whereas the highest number of outliers are shared between locations within each country (Figure 4), with Norway presenting the highest values (between 8 and 14 outliers, 21% and 36.8%, respectively) followed by the UK (7 outliers, 20%) (Table 2). The number and proportion of shared outliers between locations of different countries were generally lower, ranging from 2 (6.9%) between Lökholmen (Sweden) and Syltøyra (Norway) to 7 (14.6%) between Anglesey North (UK) and Hummelsund (Norway; Table 2).

In total, 147 outliers ($p > .95$) were detected across locations; this is combining the outlier loci that were detected between habitats within each location. The NJ tree based on these 147 outliers shows that the populations group first by habitat and only then by country within each habitat (Figure 5a). The two Ursholmen populations cluster together within the “sheltered” clade confirming the absence of a moderately exposed site within this location in our dataset. Consequently, this site was excluded from the discussion on the main patterns revealed by this study, except when otherwise specified.

The STRUCTURE analyses for outlier loci rendered $K = 2$ as the most likely number of genetic clusters regardless of the geographic scale (global, regional and local; see Methods and Figure 6a). The results are similar across all three levels (correlation between the individuals' membership ($r > 0.92$, $p < 2.2e-16$), showing that the two genetic clusters roughly coincide with the two habitats (with the exception of Ursholmen, ($r < -0.02$, $p > .76$ between local memberships and the two other levels). However, some individuals present a genetic composition that is typical of the opposite habitat where they were sampled, whereas others are genetically admixed

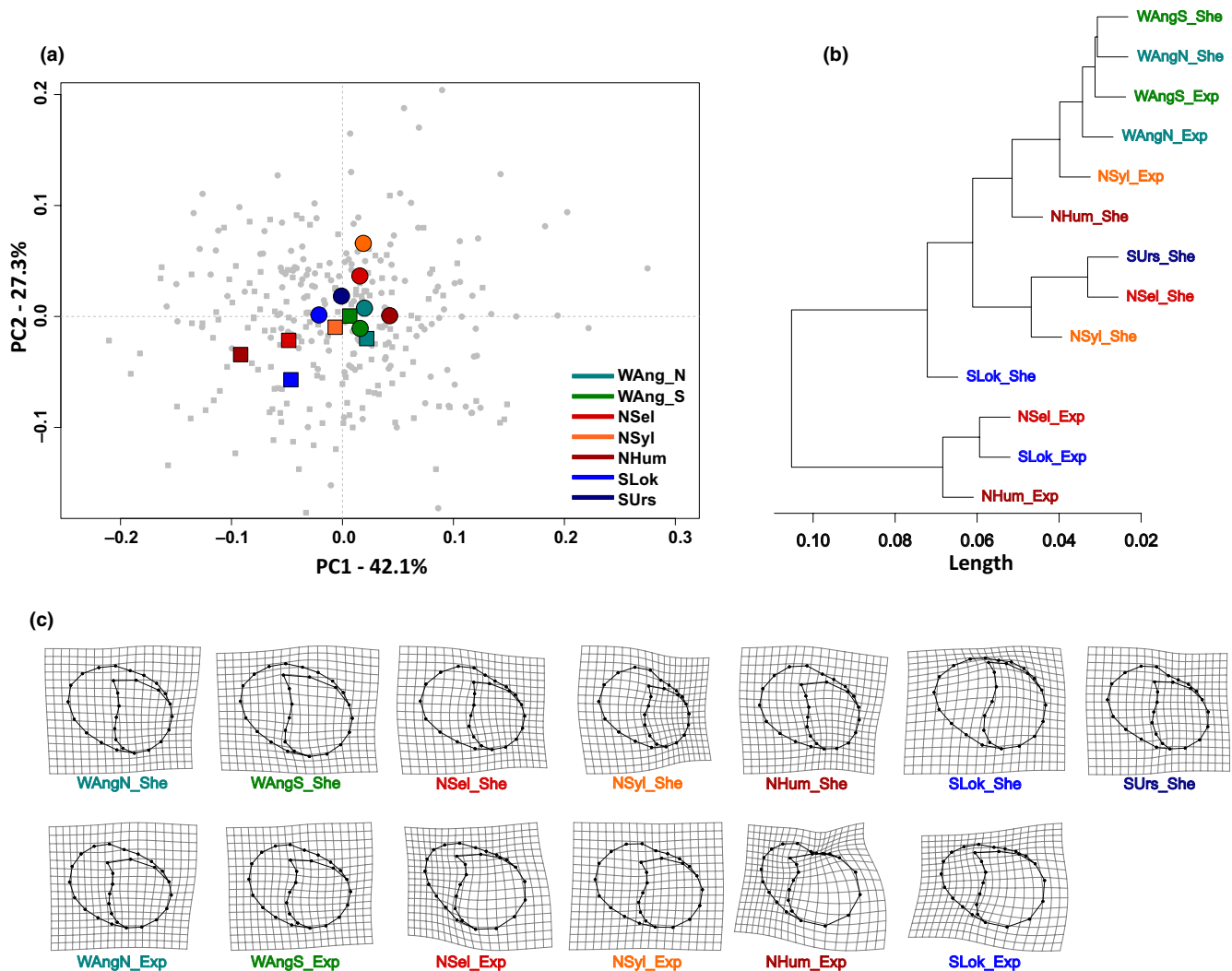


FIGURE 3 Results of the geometric morphometric (GM) analyses of shell shape. (a) Mean shape of individuals of each habitat and location based on the two first principal components of the PCA, where mean values are indicated by coloured symbols, whereas individuals are represented in grey; square and circle symbols represent the moderately exposed and sheltered habitats, respectively. (b) UPGMA dendrogram based on the Procrustes shape distances. (c) Deformation grids depicting the average shape of individuals of each habitat and location compared to the global mean. Mean shapes were magnified $\times 2$ to improve visualization. For Ursholmen, only samples of the sheltered habitat were included (see main text). Population codes are the same as in Table S1 and Figure S1

between the two clusters, suggesting migration and interbreeding between individuals from the two habitats.

3.3 | Genetic substructure based on nonoutlier loci

In contrast, the NJ tree for nonoutlier loci (380 loci, combining the nonoutlier loci detected individually in each pairwise comparison) (Figure 5b) shows that the populations primarily group by country and only then by location. The STRUCTURE results for $K = 2$ based on nonoutliers also contrasted with those obtained based on outliers (Figure 6b). Instead of the two main clusters representing the habitats where the individuals were found, the two main clusters separate UK from Scandinavian populations (for $K = 3$) and only for $K = 4$ a split between Norwegian and Swedish populations is observed. A

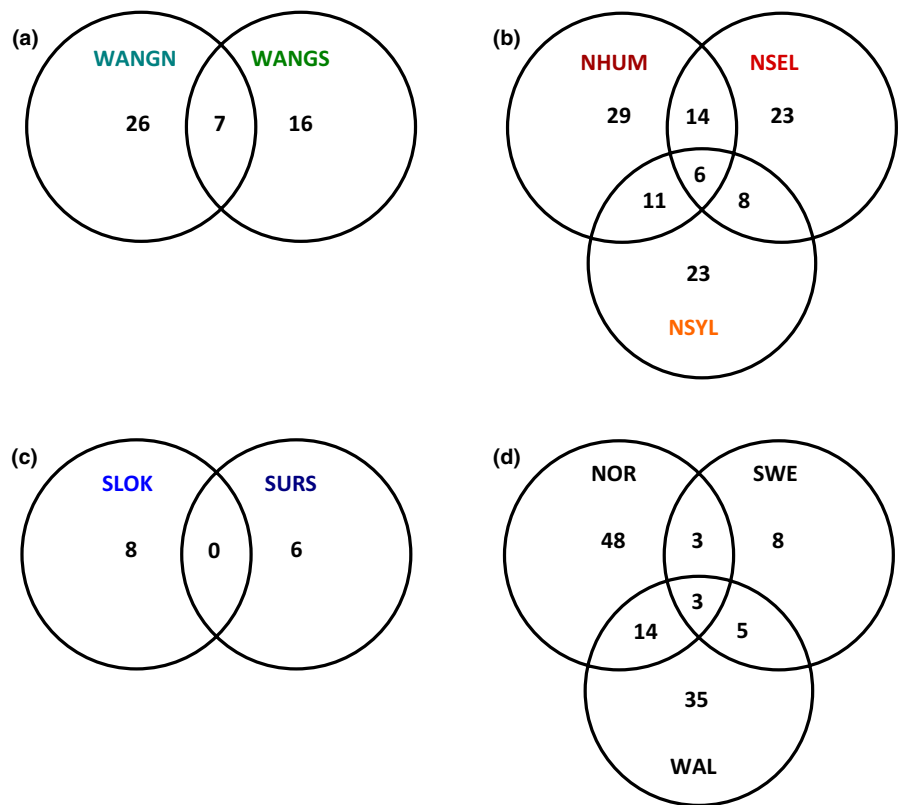
similar substructure was observed for $K = 7$ (Figure S3), the most probable number of clusters according to the Evanno method (not shown).

Mean pairwise F_{ST} values show the lowest differentiation between populations from Norway and Sweden, followed by populations from Norway and UK and then by populations from Sweden and UK, in agreement with the geographic distance between countries (Figure S4A). The genetic differentiation between locations is relatively higher in UK (Figure S4B), followed by Sweden and Norway. Within each country, the populations from sheltered habitats showed higher differentiation than the exposed ones, except in Norway where F_{ST} between Syltøyana and Seløyana is close to zero in both sheltered and moderately exposed habitats (Figure S4D). Pairwise F_{ST} between populations living in different habitats within each location was zero, whereas this was generally not the case between populations from the

TABLE 1 Results of the outlier detection analysis performed with BAYESCAN and DFDIST at different significance levels ($p > .99$, $p > .95$) for each location

Location	F_{ST} overall	BAYESCAN	DFDIST N outlier 99	F_{ST} outlier 99	DFDIST N outlier 95	F_{ST} outlier 95	DFDIST N nonoutlier	F_{ST} nonoutlier
WAngN	0.0469	2 (0.4)	26 (4.7)	0.1886	53 (9.5)	0.1488	437 (78.6)	0
WAngS	0.0369	5 (0.9)	16 (2.8)	0.1699	36 (6.4)	0.1114	454 (80.2)	0
NSel	0.0557	8 (1.5)	23 (4.2)	0.2304	41 (7.5)	0.1581	434 (79.2)	0
NSyl	0.0363	2 (0.4)	23 (4.1)	0.1425	42 (7.5)	0.1063	442 (79.2)	0
NHum	0.0805	9 (1.7)	29 (5.6)	0.2922	48 (9.2)	0.2079	422 (81.0)	0
SLok	0.0208	0	8 (1.6)	0.0895	31 (6.3)	0.0682	420 (84.7)	0
SUrs	0.0109	0	6 (1.2)	0.0364	27 (5.3)	0.0410	414 (80.9)	0

Note: The number of outlier loci and percentage (between brackets) over the total number of loci analysed is shown. The number and percentage of nonoutlier (putatively neutral) loci (see Methods) is also shown. Between ecotype F_{ST} values were calculated in AFLP-SURV overall loci, with all outlier 99 ($p > .99$), all outlier 95 ($p > .95$), and all nonoutlier ($p < .80$).

FIGURE 4 Number of shared DFDIST outliers ($p > .99$) between locations for Wales, United Kingdom (a), Norway (b), Sweden (c); and between countries (taking into account all the outliers across locations within a country) (d), except Sweden for which only the outliers detected in Lökholmen were considered (see main text). The total number of outliers within locations is represented inside the circles (including the shared ones)**TABLE 2** Number of shared DFDIST outliers ($p > .99$, below diagonal; $p > .95$, above diagonal) between locations

Location	WAngN	WAngS	NSel	NSyl	NHum	SLok	SUrs
WAngN		16 (22.0)	11 (13.2)	11 (13.1)	13 (14.8)	10 (13.5)	9 (12.7)
WAngS	7 (20.0)		14 (22.2)	11 (16.4)	16 (23.5)	10 (17.5)	2 (3.3)
NSel	6 (13.9)	5 (14.7)		17 (25.7)	26 (41.3)	11 (18.0)	3 (4.6)
NSyl	4 (8.9)	3 (8.3)	8 (21.0)		20 (30.0)	9 (14.1)	3 (4.5)
NHum	7 (14.6)	5 (12.5)	14 (36.8)	11 (26.8)		10 (14.5)	6 (8.7)
SLok	3 (9.7)	2 (9.1)	2 (6.9)	3 (10.7)	3 (8.8)		5 (9.4)
SUrs	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	

Note: In brackets, the percentage of shared outliers from the total number of outliers in the pairwise comparison.

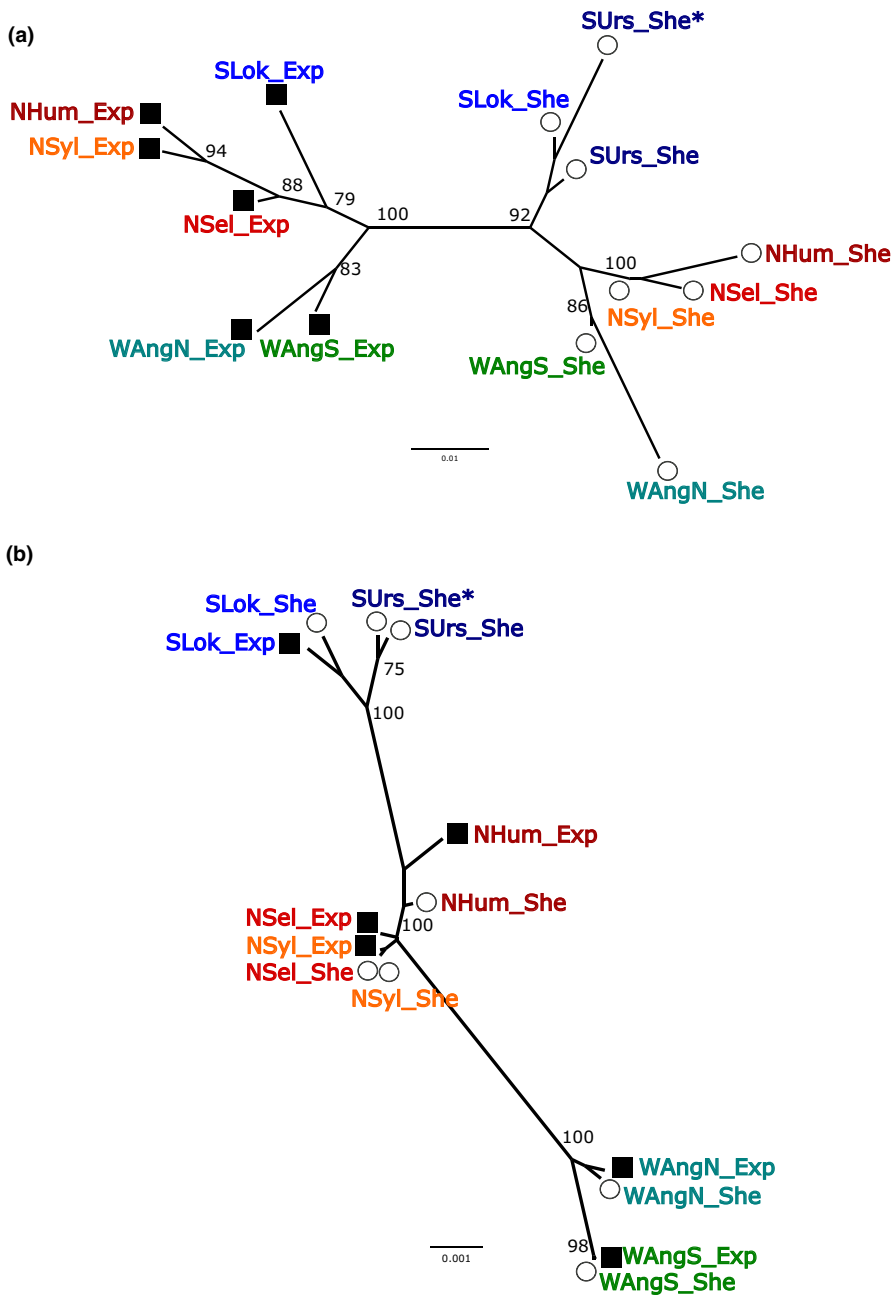


FIGURE 5 Neighbour-Joining trees based on Nei's genetic distance calculated with AFLP-SURV in two sets of loci: (a) outliers 95 ($p > .95$, DFDIST) (147 loci) and (b) nonoutliers ($p < .80$) (380 loci), in both cases combining the loci from all comparisons (see main text). Population codes are the same as in Table S1 and Figure S1. Dark squares represent populations from exposed sites, whereas white circles represent populations from sheltered sites. Only bootstrap support values above 70 are shown

same habitat among locations, apart from moderately exposed locations from Norway (Figure S4 and Table S3).

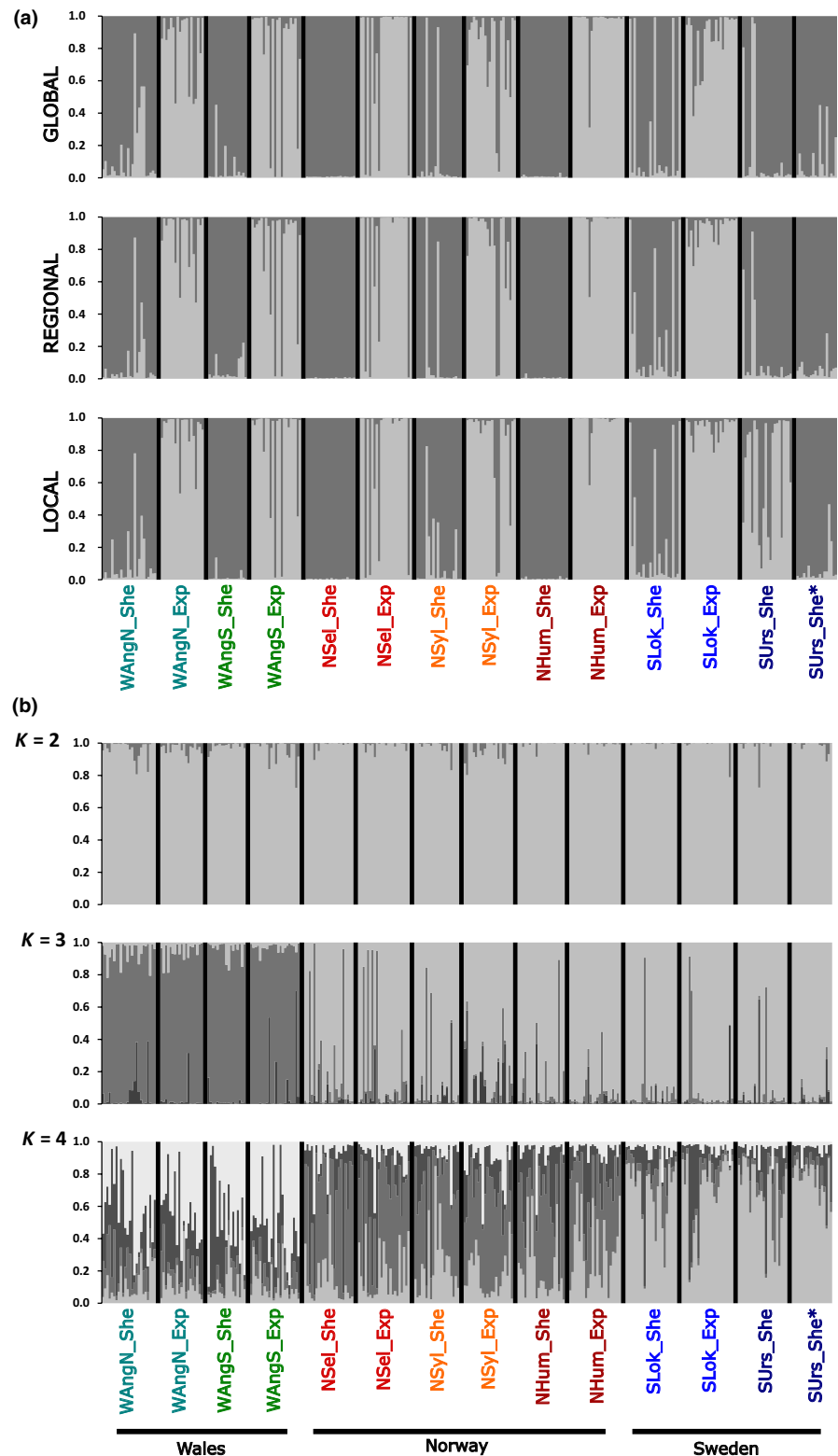
Using a $PP \geq 0.90$ criterion to classify individuals as one of two ecotypes, 49 individuals over a total of 83 analysed for both genetics and shape show concordant classification between the two types of information, with the majority (96%, $N = 47$) of these samples assigned to the cluster/form typical of the habitat where they were sampled (Table S4). The proportion of concordance was similar between sheltered and moderately exposed habitats (59% and 55%, respectively), and although this varies substantially between sites, we refrain to draw any conclusions given the small sample sizes per site (from 1 to 11, Table S4). Some individuals that are genetically classified as pure for the typical sheltered cluster show a typical shape of the exposed habitat ($N = 9$), and vice versa ($N = 5$).

However, most of these individuals ($N = 9$) belong to the genetic cluster typical of the habitat where they were sampled (Figure S5, top left and bottom right). Finally, genetically admixed individuals ($N = 14$) present a wide range of shell shape, with the majority ($N = 9$) showing a morphology typical of moderately exposed habitats.

4 | DISCUSSION

Studies of systems like *L. fabalis*, where different ecotypes coexist across a species' distribution range, offer important insights about the genetic variation underlying traits involved in local adaptation and ecological speciation. Combining an AFLP genome scan for

FIGURE 6 STRUCTURE plots ($2 \leq K \leq 4$) for: (a) three hierarchical levels for $K = 2$ (global, regional, and local; see Methods) using outlier loci ($p > .95$, DFDIST) (147 loci) and (b) nonoutliers ($p < .80$) (380 loci), in both cases combining the loci from all comparisons (see main text). Population codes are the same as in Table S1 and Figure S1. Each cluster is represented by a different grey shade. For the global analysis, shades are comparable across all countries and locations, whereas for the regional analysis shades are comparable only within countries. Shades are not comparable among locations in the local analysis



outlier loci with geometric morphometric analysis, we made the first attempt to detect genetic variation associated with divergent ecotypes in terms of wave-exposure and/or related ecological factors (environmental or biological) and to evaluate the level of sharing of these outlier loci between ecotypes across populations from the UK, Sweden and Norway.

4.1 | The role of natural selection on phenotypic divergence

In agreement with previous studies (Kempainen et al., 2005, 2011; Tatarenkov & Johannesson, 1998), mean shell size was larger in moderately exposed than in sheltered sites (Figure 2a). The remarkable

influence of habitat type in moulding size variation is evident in the size-based UPGMA dendrograms where moderately exposed sites (with a single exception) consistently cluster together, irrespective of their geographic origin (Figure 2b). Although shell shape variation was also influenced by geography, divergence between populations from the two contrasting habitats was significant across locations (Figure 3a). Thus, despite local-specific phenotypic effects, we found consistent divergence in shell morphology between *L. fabalis* populations at the extremes of similar environmental transitions across localities from the same or different European regions. Although this could not be assessed in Ursholmen, observations from a moderately exposed site recently visited show that this pattern holds for this island too, at least for size (R. Faria, personal observation).

The patterns of phenotypic and genetic variation can be explained by both genetic and environmental factors. Although a plastic component cannot be excluded, the association between size/growth and genetic variation found by Tatarenkov and Johannesson (1998) in individuals originally from the same intermediately exposed habitat suggests an important heritable component. This is in agreement with variation in shell morphology being largely genetically inherited in other species within the genus, such as *L. saxatilis* (Conde-Padín et al., 2009; Conde-Padín et al., 2007; Galindo et al., 2019; Hollander & Butlin, 2010; Johannesson & Johannesson, 1996; Johannesson et al., 1997) and *L. subrotundata* (Boulding & Hay, 1993; Kyle & Boulding, 1998). Thus, shell divergence between habitats across multiple populations of *L. fabalis* strongly suggests local adaptation to different levels of wave exposure and/or related ecological factors. For example, a larger shell aperture is observed in exposed sites when compared with sheltered sites in *L. fabalis* (Figure 3c), which agrees with earlier findings in *L. saxatilis* (Johannesson et al., 2010). In contrast, *L. fabalis* from central and northern Europe are larger in moderately exposed habitats than in sheltered habitats (Reimchen, 1981; Tatarenkov & Johannesson, 1998), whereas the opposite is true in *L. saxatilis* (Johannesson et al., 2010). However, both the sheltered and moderately exposed habitats of *L. fabalis* correspond to the "crab" sites of *L. saxatilis*, whereas the "wave" sites of *L. saxatilis* are far more exposed and crab free.

A previous ecological study of the two *L. fabalis* ecotypes suggested that morphological differences are due to a complex interaction between wave exposure and the algae canopy inhabited by the snails, which provides shelter against crabs living in the boulders below (Kemppainen et al., 2005). According to this hypothesis, individuals with a larger and thicker shell would be favoured against predators in moderately exposed habitats where dislodgment is more likely. The shape patterns observed here are consistent with this hypothesis as the large shell aperture characteristic of individuals from moderately exposed habitats can accommodate a larger foot and protect them against dislodgement. In this sense, the moderately exposed ecotype seems to be adjusted for both higher predation (large size) and avoiding dislodgement (large aperture relative to body size), in contrast to *L. saxatilis* where the exposed ecotype does not need protection against crab predation and is only adapted to fit into crevices and cling tightly to the rocks to avoid wave dislodgement (reviewed in Johannesson et al., 2010).

Strikingly, the moderately exposed population of Syltøyna (Norway) groups with the sheltered populations when considering both size and shape (Figures 2b and 3b). This suggests that selection related with wave action is weaker in this site, which is supported by our observations in the field. Independently of the causes, this highlights that besides general habitat-related differences there are also important location-specific effects on shell morphology (Table S2).

4.2 | Habitat-related genetic differentiation

As commonly found in other studies (reviewed by Ravinet et al., 2017), our results reveal heterogeneous differentiation between ecotypes, with a relatively small proportion of loci showing high levels of differentiation likely resisting the substantial gene flow that seems to be eroding differentiation to very low levels in the rest of the genome here assessed. However, an important caveat of genome scans is that the number (and proportion) of discovered outliers depends on how stringent is the cut-off used to detect them (Faria et al., 2014). Here, we used different methods, including BAYESCAN that is known to be conservative (Pérez-Figueroa et al., 2010), as well as different stringency levels to detect outliers. For instance, when considering the less stringent criterion and method (DFDIST, $p > .95$), we detected ~7% of outliers across locations, which is similar to that detected between other pairs of divergent ecotypes in a wide range of taxa (reviewed in Nosil et al., 2009), including the Crab and Wave ecotypes of *L. saxatilis* (Butlin et al., 2014). Nonetheless, as with any genome scan, these outliers must be seen as a list of potential candidate loci influenced by divergent selection that need further confirmation by alternative approaches, as suggested by Ravinet et al. (2017).

Outlier loci identified by means of genome scans could also result from factors not related with local adaptation, such as background selection (Cruickshank & Hahn, 2014; Ravinet et al., 2017). However, given that F_{ST} between habitats based on nonoutliers is zero in all locations, it is very unlikely that background selection would be strong enough to originate peaks of F_{ST} under such substantial levels of gene flow. An important limitation of using AFLPs is that we cannot directly assess the degree of physical linkage between loci (Stapley et al., 2010). Thus, we do not know if these outliers are clustered within a single genomic region or are widespread across the genome, limiting our power to further interpret the differentiation heterogeneity. However, the consistent patterns of differentiation between habitats at different geographical scales (global, regional and local) suggest that the outliers we detected are indeed influenced by the same axis of selection across locations. The proportion of outliers and associated genetic differentiation vary across locations, with the highest F_{ST} and number of outliers in Hummelsund (Norway), followed by intermediate values in UK and lower differentiation in Sweden. Given the absence of significant genetic differentiation based on nonoutlier loci in all locations (Sotelo et al., 2020; this study), these patterns are compatible with differences in the strength of selection, which can result in geographic heterogeneity

under migration–selection equilibria. Nevertheless, future studies of hybrid zones based on cline analysis of loci across the genome, like the one implemented by Westram et al. (2018), will be important to confirm the role of divergent selection in this system.

4.3 | Overlap between outliers across locations

A small number of outliers (3, $p > .99$) were shared among all countries but these estimates of outlier sharing need to be interpreted with caution. Although the genome size of *L. fabalis* is currently unknown, data from closely related species (*L. obtusata* and *L. saxatilis*) suggest a genome size of 1.2–1.35 Gb (Animal Genome Size Database 2.0, <http://www.genomesize.com/>). Thus, the 746 AFLP loci we genotyped here confer relatively low resolution to assess both the number of outliers and the amount of sharing across locations with high precision when compared, for instance, with whole-genome sequencing. However, the number of shared outliers across the three countries using the most stringent criterion is higher than what would be expected just by chance (0). Furthermore, the trend for higher outlier sharing at smaller geographic scales, as we observe here when we compare locations within and among countries, is consistent with the hypothesis of evolution in concert, where some adaptive alleles spread over populations through gene flow whereas others originated locally (Johannesson et al., 2010; Kempainen et al., 2011). Alternatively, similar patterns could also have resulted from shared standing genetic variation inherited from a common ancestral population (Westram et al., 2016). In fact, accumulated evidence from multiple systems suggests that an important component of the variation involved in repeated episodes of adaptation is relatively old and maintained as standing genetic variation (e.g. Marques et al., 2019; Roesti et al., 2015).

Ultimately, the level of outlier sharing also depends on the genomic architecture of the traits under selection, including whether they have a polygenic basis versus just a few genes of large effect involved, the genomic distribution of the underlying genes and on the recombination landscape across the genome. Indeed, genes involved in adaptation and speciation tend to cluster in regions of low recombination (Ravinet et al., 2017 and references therein). The high linkage disequilibrium between loci within these regions can lead to an increase in the amount of outliers shared across locations, even if just one locus is influenced by selection or if the selective pressures actually differ among locations (Berner & Roesti, 2017; Haenel et al., 2018; Roesti et al., 2012). This is particularly true within inverted regions, where recombination is heavily reduced in heterokaryotypes, as shown in multiple systems (Wellenreuther & Bernatchez, 2018), including *L. saxatilis* (Faria et al., 2019; Morales et al., 2019). The presence of inversions in *L. fabalis* has been previously hypothesized based on strong association between snail size, Ark genotype, and a RAPD locus genotype (Johannesson & Mikhailova, 2004; Tatarenkov & Johannesson, 1999). Since the location of the AFLP loci (anonymous markers) in the *L. fabalis* genome is unknown, a contiguous reference genome together with

a high-resolution genome scan and linkage maps will be needed to understand the genomic architecture of adaptation in this system and how it relates with the recombination landscape. In particular, the characterization of transects across the environmental gradient using targeted-capture or whole-genome sequencing, as those performed in *L. saxatilis* (Faria et al., 2019; Westram et al., 2018), will be important to inform us about the role of inversions in *L. fabalis* diversification.

The contrast between the patterns of genetic structure based on outlier loci and on putatively neutral variation is noteworthy. The clustering of populations from both ecotypes by geography, when considering nonoutlier loci, suggests parallel evolution of *L. fabalis* ecotypes (Figures 4 and 5). However, gene flow between populations from different habitats is likely high enough to generate a pattern of parallel evolution, even if they had a single origin and came into secondary contact across multiple locations (Faria et al., 2014). Thus, a modelling approach, possibly based on approximate Bayesian computation and using sequence data across the genome needs to be implemented in future studies to infer the demographic history of ecotypes and specifically test whether parallel evolution, as observed in *L. saxatilis* (Butlin et al., 2014), is more likely than a single origin of ecotypes. Nevertheless, the fact that almost half of all outliers (79 of 147 for $p > .95$) are private to one location suggests that at least some components of divergent evolution are site-specific. Consequently, even if some of the genetic variation involved in ecotype differentiation had a single origin and was shared after secondary contact, each ecotype is likely to follow its own evolutionary trajectory in each location.

4.4 | Implications for the study of speciation

Studies of local adaptation in intertidal habitats of rocky shores are key to quantify the contribution of ecological speciation to marine biodiversity (Sanford & Kelly, 2011). Overall, our results are consistent with the role of natural selection in driving divergence between *L. fabalis* ecotypes in the presence of gene flow. Previous mate choice experiments have shown that, although both males and females from the large ecotype tend to mate assortatively, males of the small ecotype can mate with both small and large ecotype females (Saltin et al., 2013). However, while mate choice is known to play an important role in reproductive isolation in *L. saxatilis* (Perini et al., 2020), whether this partial reproductive barrier is likely to result in significant reproductive isolation remains to be evaluated in natural populations for *L. fabalis*. Nevertheless, the maintenance of habitat-related phenotypic differentiation in *L. fabalis* despite high gene flow likely involves multiple loci influencing different traits, including shell thickness, growth and shape, suggesting some degree of reproductive isolation generated by extrinsic (ecological) factors. Although more detailed studies are needed to confirm and extend some of the reported results, the heterogeneous patterns of differentiation here identified, where a relatively small proportion of loci resist the homogenizing effects

of gene flow in comparison with lack of differentiation at most studied loci, is compatible with initial stages of ecological speciation (Nosil, 2012; Seehausen et al., 2014). The *L. fabalis* system thus comprises an interesting system from the marine environment where information from multiple instances of divergence across an environmental transition opens doors for further studies on how populations cope with environmental changes (Sgrò et al., 2011) and how different reproductive barriers accumulate during speciation.

ACKNOWLEDGMENTS

We would like to thank Carolina Pereira for her help in processing and extracting DNA of some samples. This study was supported by: European Regional Development Fund (FCOMP-01-0124-FEDER-014272), FCT - Foundation for Science and Technology (PTDC/BIA-EVF/113805/2009) and ASSEMBLE (grant agreement number 227799). RF was financed by FCT (SFRH/BPD/89313/2012) and by the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement number 706376; and is currently funded by. JC was funded by a PhD scholarship from FCT (PD/BD/128350/2017). GS was financed by Volkswagen Stiftung (50500776). AK was supported by an IF contract (IF/00641/2014/CP1256/CT0008) from Fundação para a Ciência e a Tecnologia (FCT, Portugal). JG was funded by a JIN project (Jóvenes Investigadores sin vinculación o con vinculación temporal, Ministerio de Ciencia, Innovación y Universidades, Spain, RTI2018-101274-J-I00).

CONFLICT OF INTEREST

The authors declare no conflicts of interest.

AUTHOR CONTRIBUTIONS

JG, PK, KJ and RF designed the study and planned the experiments. GS, MD, DC, PK, KJ and RF sampled the individuals. JG, JC and RF performed the experiments and analysed the genetic data. DC and AK analysed the morphological data. JG, JC and RF wrote the first draft. All the authors contributed to the final version.

PEER REVIEW

The peer review history for this article is available at <https://publons.com/publon/10.1111/jeb.13705>

DATA AVAILABILITY STATEMENT


Genetic and morphological data is publicly available in Dryad: <https://doi.org/10.5061/dryad.vmcvncr2>

ORCID

Juan Galindo  <https://orcid.org/0000-0002-8631-8006>

João Carvalho  <https://orcid.org/0000-0002-1728-0075>

Diana Costa  <https://orcid.org/0000-0001-6768-1240>

Antigoni Kaliontzopoulou  <https://orcid.org/0000-0002-7897-7204>

[org/0000-0002-7897-7204](https://orcid.org/0000-0002-7897-7204)

Kerstin Johannesson  <https://orcid.org/0000-0003-0176-7986>

Rui Faria  <https://orcid.org/0000-0001-6635-685X>

REFERENCES

- Adams, D., Collyer, M., & Kaliontzopoulou, A. (2019). *Geomorph: Software for geometric morphometric analyses*. R package version 3.1.0.
- Arrigo, N., Holderegger, R., & Alvarez, N. (2012). Automated scoring of AFLPs using RawGeno v 2.0, a Free R CRAN library. In: F. Pompanon, & A. Bonin (Eds.), *Data production and analysis in population genomics. methods in molecular biology (methods and protocols)*, (-) 888 pp. 155-175. Humana Press.
- Beaumont, M. A., & Nichols, R. A. (1996). Evaluating loci for use in the genetic analysis of population structure. *Proceedings of the Royal Society B-Biological Sciences*, 263, 1619-1626.
- Berner, D., & Roesti, M. (2017). Genomics of adaptive divergence with chromosome-scale heterogeneity in crossover rate. *Molecular Ecology*, 26, 6351-6369. <https://doi.org/10.1111/mec.14373>
- Boulding, E. G., & Hay, T. K. (1993). Quantitative genetics of shell form of an intertidal snail: Constraints on short-term response to selection. *Evolution*, 47, 576-592. <https://doi.org/10.1111/j.1558-5646.1993.tb02114.x>
- Butlin, R. K., Saura, M., Charrier, G., Jackson, B., André, C., Caballero, A., Coyne, J. A., Galindo, J., Grahame, J. W., Hollander, J., Kempainen, P., Martínez-Fernández, M., Panova, M., Quesada, H., Johannesson, K., & Rolán-Alvarez, E. (2014). Parallel evolution of local adaptation and reproductive isolation in the face of gene flow. *Evolution*, 68, 935-949. <https://doi.org/10.1111/evo.12329>
- Carvajal-Rodríguez, A., Conde-Padín, P., & Rolán-Alvarez, E. (2005). Decomposing shell form into size and shape by geometric morphometric methods in two sympatric ecotypes of *Littorina saxatilis*. *Journal of Molluscan Studies*, 71, 313-318. <https://doi.org/10.1093/mollus/eyi037>
- Charbit, S., Ritz, C., Philippon, G., Peyaud, V., & Kageyama, M. (2007). Numerical reconstructions of the Northern Hemisphere ice sheets through the last glacial-interglacial cycle. *Climate of the Past*, 3, 15-37. <https://doi.org/10.5194/cp-3-15-2007>
- Collyer, M. L., & Adams, D. C. (2018). RRPP: An r package for fitting linear models to high-dimensional data using residual randomization. *Methods in Ecology and Evolution*, 9, 1772-1779.
- Collyer, M., & Adams, D. (2019). *RRPP: Linear model evaluation with randomized residuals in a permutation procedure*. R package version 0.4.0.
- Conde-Padín, P., Caballero, A., & Rolán-Alvarez, E. (2009). Relative role of genetic determination and plastic response during ontogeny for shell-shape traits subjected to diversifying selection. *Evolution*, 63, 1356-1363. <https://doi.org/10.1111/j.1558-5646.2009.00636.x>
- Conde-Padín, P., Grahame, J. W., & Rolán-Alvarez, E. (2007). Detecting shape differences in species of the *Littorina saxatilis* complex by morphometric analysis. *Journal of Molluscan Studies*, 73, 147-154. <https://doi.org/10.1093/mollus/eym009>
- Connell, J. H. (1972). Community interactions on marine rocky intertidal shores. *Annual Review of Ecology and Systematics*, 3, 169-192. <https://doi.org/10.1146/annurev.es.03.110172.001125>
- Costa, D., Sotelo, G., Kaliontzopoulou, A., Carvalho, J., Butlin, R., Hollander, J., Faria, R. (2020). Hybridization patterns between two marine snails, *Littorina fabalis* and *L. obtusata*. *Ecology and Evolution*, 10, 1158-1179.
- Coyer, J. A., Hoarau, G., Costa, J. F., Hogerdijk, B., Serrão, E. A., Billard, E., Valero, M., Pearson, G. A., & Olsen, J. L. (2011). Evolution and diversification within the intertidal brown macroalgae *Fucus spiralis*/*F. vesiculosus* species complex in the North Atlantic. *Molecular Phylogenetics and Evolution*, 58, 283-296. <https://doi.org/10.1016/j.ympev.2010.11.015>
- Cruickshank, T. E., & Hahn, M. W. (2014). Reanalysis suggests that genomic islands of speciation are due to reduced diversity, not reduced gene flow. *Molecular Ecology*, 23, 3133-3157.
- Denny, M., & Wetthey, D. (2001). Physical processes that generate patterns in marine communities. In Bertness, M. D.; Gaines, S. D. & Hay,

- M. E. (eds.), *Marine Community Ecology*, pp. 3–37. Sunderland: Sinauer Associates.
- Elmer, K. R., & Meyer, A. (2011). Adaptation in the age of ecological genomics: Insights from parallelism and convergence. *Trends in Ecology & Evolution*, *26*, 298–306. <https://doi.org/10.1016/j.tree.2011.02.008>
- Evanno, G., Regnaut, S., & Goudet, J. (2005). Detecting the number of clusters of individuals using the software STRUCTURE: A simulation study. *Molecular Ecology*, *14*, 2611–2620. <https://doi.org/10.1111/j.1365-294X.2005.02553.x>
- Faria, R., Chaube, P., Morales, H. E., Larsson, T., Lemmon, A. R., Lemmon, E. M., Rafajlović, M., Panova, M., Ravinet, M., Johannesson, K., & Westram, A. M. (2019). Multiple chromosomal rearrangements in a hybrid zone between *Littorina saxatilis* ecotypes. *Molecular Ecology*, *28*, 1375–1393.
- Faria, R., Renaut, S., Galindo, J., Pinho, C., Melo-Ferreira, J., Melo, M., Jones, F., Salzburger, W., Schluter, D., & Butlin, R. (2014). Advances in ecological speciation: An integrative approach. *Molecular Ecology*, *23*, 513–521. <https://doi.org/10.1111/mec.12616>
- Felsenstein, J. (2013). *PHYLP (phylogeny inference package)*. Department of Genome Sciences, University of Washington, Seattle, Washington.
- Foll, M., & Gaggiotti, O. (2008). A genome-scan method to identify selected loci appropriate for both dominant and codominant markers: A Bayesian perspective. *Genetics*, *180*, 977–993. <https://doi.org/10.1534/genetics.108.092221>
- Galindo, J., Cacheda, D., Caballero, A., & Rolán-Alvarez, E. (2019). Untangling the contribution of genetic and environmental effects to shell differentiation across an environmental cline in a marine snail. *Journal of Experimental Marine Biology and Ecology*, *513*, 27–34. <https://doi.org/10.1016/j.jembe.2019.02.004>
- Galindo, J., & Grahame, J. W. (2014). Ecological Speciation and the Intertidal Snail *Littorina saxatilis*. *Advances in Ecology*. 2014 1-9. <https://doi.org/10.1155/2014/239251>
- Galindo, J., Martínez-Fernández, M., Rodríguez-Ramilo, S. T., & Rolán-Alvarez, E. (2013). The role of local ecology during hybridization at the initial stages of ecological speciation in a marine snail. *Journal of Evolutionary Biology*, *26*, 1472–1487. <https://doi.org/10.1111/jeb.12152>
- Galindo, J., Morán, P., & Rolán-Alvarez, E. (2009). Comparing geographical genetic differentiation between candidate and noncandidate loci for adaptation strengthens support for parallel ecological divergence in the marine snail *Littorina saxatilis*. *Molecular Ecology*, *18*, 919–930.
- Gorelick, N., Hancher, M., Dixon, M., Ilyushchenko, S., Thau, D., & Moore, R. (2017). Google Earth Engine: Planetary-scale geospatial analysis for everyone. *Remote Sensing of Environment*, *202*, 18–27. <https://doi.org/10.1016/j.rse.2017.06.031>
- Grahame, J. W., Wilding, C. S., & Butlin, R. K. (2006). Adaptation to a steep environmental gradient and an associated barrier to gene exchange in *Littorina saxatilis*. *Evolution*, *60*, 268–278. <https://doi.org/10.1111/j.0014-3820.2006.tb01105.x>
- Haenel, Q., Laurentino, T. G., Roesti, M., & Berner, D. (2018). Meta-analysis of chromosome-scale crossover rate variation in eukaryotes and its significance to evolutionary genomics. *Molecular Ecology*, *27*, 2477–2497. <https://doi.org/10.1111/mec.14699>
- Helmuth, B., & Denny, M. W. (2003). Predicting wave exposure in the rocky intertidal zone: Do bigger waves always lead to larger forces? *Limnology and Oceanography*, *48*, 1338–1345.
- Hollander, J., & Butlin, R. K. (2010). The adaptive value of phenotypic plasticity in two ecotypes of a marine gastropod. *BMC Evolutionary Biology*, *10*, 333. <https://doi.org/10.1186/1471-2148-10-333>
- Hollander, J., Galindo, J., & Butlin, R. K. (2015). Selection on outlier loci and their association with adaptive phenotypes in *Littorina saxatilis* contact zones. *Journal of Evolutionary Biology*, *28*, 328–337.
- Johannesson, B., & Johannesson, K. (1996). Population differences in behaviour and morphology in the snail *Littorina saxatilis*: Phenotypic plasticity or genetic differentiation? *Journal of Zoology*, *240*, 475–493.
- Johannesson, K. (2001). Parallel speciation: A key to sympatric divergence. *Trends in Ecology & Evolution*, *16*, 148–153. [https://doi.org/10.1016/S0169-5347\(00\)02078-4](https://doi.org/10.1016/S0169-5347(00)02078-4)
- Johannesson, K., & Mikhailova, N. (2004). Habitat-related genetic substructuring in a marine snail (*Littorina fabalis*) involving a tight link between an allozyme and a DNA locus. *Biological Journal of the Linnean Society*, *81*, 301–306. <https://doi.org/10.1111/j.1095-8312.2003.00288.x>
- Johannesson, K., Panova, M., Kempainen, P., André, C., Rolán-Alvarez, E., & Butlin, R. K. (2010). Repeated evolution of reproductive isolation in a marine snail: Unveiling mechanisms of speciation. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, *365*, 1735–1747. <https://doi.org/10.1098/rstb.2009.0256>
- Johannesson, K., Rolán-Alvarez, E., & Erlandsson, J. (1997). Growth rate differences between upper and lower shore ecotypes of the marine snail *Littorina saxatilis* (Olivi) (Gastropoda). *Biological Journal of the Linnean Society*, *61*, 267–279. <https://doi.org/10.1111/j.1095-8312.1997.tb01790.x>
- Kempainen, P., Lindskog, T., Butlin, R., & Johannesson, K. (2011). Intron sequences of arginine kinase in an intertidal snail suggest an ecotype-specific selective sweep and a gene duplication. *Heredity*, *106*, 808–816.
- Kempainen, P., Panova, M., Hollander, J., & Johannesson, K. (2009). Complete lack of mitochondrial divergence between two species of NE Atlantic marine intertidal gastropods. *Journal of Evolutionary Biology*, *22*, 2000–2011. <https://doi.org/10.1111/j.1420-9101.2009.01810.x>
- Kempainen, P., Van Nes, S., Ceder, C., & Johannesson, K. (2005). Refuge function of marine algae complicates selection in an intertidal snail. *Oecologia*, *143*, 402–411. <https://doi.org/10.1007/s00442-004-1819-5>
- Kess, T., Galindo, J., & Boulding, E. G. (2018). Genomic divergence between Spanish *Littorina saxatilis* ecotypes unravels limited admixture and extensive parallelism associated with population history. *Ecology and Evolution*, *8*, 8311–8327.
- Kyle, C. J., & Boulding, E. G. (1998). Molecular genetic evidence for parallel evolution in a marine gastropod, *Littorina subrotundata*. *Proceedings of the Royal Society B-Biological Sciences*, *265*, 303–308.
- Le Pennec, G., Butlin, R. K., Jonsson, P. R., Larsson, A. I., Lindborg, J., Bergström, E., Westram, A. M., & Johannesson, K. (2017). Adaptation to dislodgement risk on waveswept rocky shores in the snail *Littorina saxatilis*. *PLoS One*, *12*, e0186901. <https://doi.org/10.1371/journal.pone.0186901>
- Little, C., & Kitching, J. A. (1996). *The biology of rocky shores*. Oxford University Press.
- Lynch, M., & Milligan, B. G. (1994). Analysis of population genetic structure with RAPD markers. *Molecular Ecology*, *3*, 91–99. <https://doi.org/10.1111/j.1365-294X.1994.tb00109.x>
- Marques, D. A., Meier, J. I., & Seehausen, O. (2019). A combinatorial view on speciation and adaptive radiation. *Trends in Ecology & Evolution*, *34*(6), 531–544. <https://doi.org/10.1016/j.tree.2019.02.008>
- Morales, H. E., Faria, R., Johannesson, K., Larsson, T., Panova, M., Westram, A. M., & Butlin, R. K. (2019). Genomic architecture of parallel ecological divergence: beyond a single environmental contrast. *Science Advances*, *5*, eaav9663. <https://doi.org/10.1126/sciadv.aav9663>
- Nosil, P. (2012). *Ecological speciation*. Oxford University Press.
- Nosil, P., Funk, D. J., & Ortiz-Barrientos, D. (2009). Divergent selection and heterogeneous genomic divergence. *Molecular Ecology*, *18*, 375–402. <https://doi.org/10.1111/j.1365-294X.2008.03946.x>
- Paine, R. T., Fenchel, T., & Kinne, O. (1994). *Marine rocky shores and community ecology: An experimentalist's perspective*. Ecology Institute.

- Pérez-Figueroa, A., García-Pereira, M. J., Saura, M., Rolán-Alvarez, E., & Caballero, A. (2010). Comparing three different methods to detect selective loci using dominant markers. *Journal of Evolutionary Biology*, 23, 2267–2276. <https://doi.org/10.1111/j.1420-9101.2010.02093.x>
- Perini, S., Rafajlović, M., Westram, A. M., Johannesson, K., & Butlin, R. K. (2020). Assortative mating, sexual selection and their consequences for gene flow in *Littorina*. *Evolution*, <https://doi.org/10.1111/evo.14027>
- Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using multilocus genotype data. *Genetics*, 155, 945–959.
- R Core Team (2018). *R: A language and environment for statistical computing*.
- R Core Team (2019). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing. Retrieved from: <https://www.R-project.org/>
- Raffaelli, D., & Hawkins, S. J. (1996). *Intertidal Ecology*, Chapman & Hall, London.
- Ravinet, M., Faria, R., Butlin, R. K., Galindo, J., Bierne, N., Rafajlović, M., Noor, M. A. F., Mehlig, B., & Westram, A. M. (2017). Interpreting the genomic landscape of speciation: A road map for finding barriers to gene flow. *Journal of Evolutionary Biology*, 30, 1450–1477. <https://doi.org/10.1111/jeb.13047>
- Reid, D. G. (1996). *Systematics and evolution of Littorina*. London: The Ray Society.
- Reimchen, T. E. (1981). Microgeographical variation in *Littorina mariae* Sacchi & Rastelli and a taxonomic consideration. *Journal of Conchology*, 30, 341–350.
- Roesti, M., Hendry, A. P., Salzburger, W., & Berner, D. (2012). Genome divergence during evolutionary diversification as revealed in replicate lake–stream stickleback population pairs. *Molecular Ecology*, 21, 2852–2862.
- Roesti, M., Kueng, B., Moser, D., & Berner, D. (2015). The genomics of ecological vicariance in threespine stickleback fish. *Nature Communications*, 6, 8767. <https://doi.org/10.1038/ncomms9767>
- Rohlf, F. J. (2006). *Tps series*. : Department of Ecology and Evolution, State University of New York.
- Rohlf, F. J., & Slice, D. (1990). Extensions of the procrustes method for the optimal superimposition of landmarks. *Systematic Zoology*, 39, 40–59. <https://doi.org/10.2307/2992207>
- Rolán-Alvarez, E., Austin, C. J., & Boulding, E. G. (2015). The contribution of the genus *Littorina* to the field of evolutionary ecology. *Oceanography and Marine Biology: an Annual Review*, 53, 157–214.
- Saltin, S. H., Schade, H., & Johannesson, K. (2013). Preference of males for large females causes a partial mating barrier between a large and a small ecotype of *Littorina fabalis* (W. Turton, 1825). *Journal of Molluscan Studies*, 79, 128–132.
- Sanford, E., & Kelly, M. W. (2011). Local adaptation in marine invertebrates. *Annual Review of Marine Science*, 3, 509–535. <https://doi.org/10.1146/annurev-marine-120709-142756>
- Schluter, D. (2000). *The ecology of adaptive radiation*. Oxford University Press.
- Schluter, D., & Conte, G. L. (2009). Genetics and ecological speciation. *Proceedings of the National Academy of Sciences USA*, 106(Suppl), 9955–9962. <https://doi.org/10.1073/pnas.0901264106>
- Seehausen, O., Butlin, R. K., Keller, I., Wagner, C. E., Boughman, J. W., Hohenlohe, P. A., Peichel, C. L., Saetre, G.-P., Bank, C., Brännström, Å., Brelsford, A., Clarkson, C. S., Eroukhanoff, F., Feder, J. L., Fischer, M. C., Foote, A. D., Franchini, P., Jiggins, C. D., Jones, F. C., ... Widmer, A. (2014). Genomics and the origin of species. *Nature Reviews Genetics*, 15(3), 176–192. <https://doi.org/10.1038/nrg3644>
- Seeley, R. H. (1986). Intense natural selection caused a rapid morphological transition in a living marine snail. *Proceedings of the National Academy of Sciences USA*, 83, 6897–6901. <https://doi.org/10.1073/pnas.83.18.6897>
- Sgrò, C. M., Lowe, A. J., & Hoffmann, A. A. (2011). Building evolutionary resilience for conserving biodiversity under climate change. *Evolutionary Applications*, 4, 326–337. <https://doi.org/10.1111/j.1752-4571.2010.00157.x>
- Smadja, C. M., & Butlin, R. K. (2011). A framework for comparing processes of speciation in the presence of gene flow. *Molecular Ecology*, 20, 5123–5140. <https://doi.org/10.1111/j.1365-294X.2011.05350.x>
- Sokal, R. R., & Rohlf, F. J. (1995). *Biometry. The principles and practice of statistics in biological research*. W. H. Freeman and Co.
- Sotelo, G., Duvetorp, M., Costa, D., Panova, M., Johannesson, K., & Faria, R. (2020). Phylogeographic history of flat periwinkles, *Littorina fabalis* and *L. obtusata*. *BMC Evolutionary Biology*, 20, 23.
- Southward, A. J. (1957). On the behaviour of barnacles: III. Further observations on the influence of temperature and age on cirral activity. *Journal of the Marine Biological Association of the United Kingdom*, 36, 323–334. <https://doi.org/10.1017/S0025315400016830>
- Stapley, J., Reger, J., Feulner, P. G. D., Smadja, C., Galindo, J., Ekblom, R., Bennison, C., Ball, A. D., Beckerman, A. P., & Slate, J. (2010). Adaptation Genomics: The next generation. *Trends in Ecology & Evolution*, 25, 705–712. <https://doi.org/10.1016/j.tree.2010.09.002>
- Tatarenkov, A., & Johannesson, K. (1994). Habitat related allozyme variation on a microgeographic-scale in the marine snail *Littorina mariae* (Prosobranchia, Littorinacea). *Biological Journal of the Linnean Society*, 53, 105–125. <https://doi.org/10.1006/bijl.1994.1063>
- Tatarenkov, A., & Johannesson, K. (1998). Evidence of a reproductive barrier between two forms of the marine periwinkle *Littorina fabalis* (Gastropoda). *Biological Journal of the Linnean Society*, 63, 349–365. <https://doi.org/10.1111/j.1095-8312.1998.tb01522.x>
- Tatarenkov, A., & Johannesson, K. (1999). Micro- and macrogeographic allozyme variation in *Littorina fabalis*; do sheltered and exposed forms hybridize? *Biological Journal of the Linnean Society*, 67, 199–212. <https://doi.org/10.1111/j.1095-8312.1999.tb01861.x>
- Tomaneck, L., & Helmuth, B. (2002). Physiological ecology of rocky intertidal organisms: A synergy of concepts. *Integrative and Comparative Biology*, 42, 771–775. <https://doi.org/10.1093/icb/42.4.771>
- Vekemans, X., Beauwens, T., Lemaire, M., & Roldán-Ruiz, I. (2002). Data from amplified fragment length polymorphism (AFLP) markers show indication of size homoplasy and of a relationship between degree of homoplasy and fragment size. *Molecular Ecology*, 11, 139–151. <https://doi.org/10.1046/j.0962-1083.2001.01415.x>
- Wellenreuther, M., & Bernatchez, L. (2018). Eco-evolutionary genomics of chromosomal inversions. *Trends in Ecology & Evolution*, 33, 427–440. <https://doi.org/10.1016/j.tree.2018.04.002>
- Westram, A. M., Panova, M., Galindo, J., & Butlin, R. K. (2016). Targeted resequencing reveals geographical patterns of differentiation for loci implicated in parallel evolution. *Molecular Ecology*, 25, 3169–3186. <https://doi.org/10.1111/mec.13640>
- Westram, A. M., Rafajlović, M., Chaube, P., Faria, R., Larsson, T., Panova, M., Ravinet, M., Blomberg, A., Mehlig, B., Johannesson, K., & Butlin, R. (2018). Clines on the seashore: The genomic architecture underlying rapid divergence in the face of gene flow. *Evolution Letters*, 2, 297–309. <https://doi.org/10.1002/evl3.74>
- Whitlock, R., Hipperson, H., Mannarelli, M., Butlin, R. K., & Burke, T. (2008). An objective, rapid and reproducible method for scoring AFLP peak-height data that minimizes genotyping error. *Molecular Ecology Resources*, 8, 725–735.
- Wilding, C. S., Butlin, R. K., & Grahame, J. (2001). Differential gene exchange between parapatric morphs of *Littorina saxatilis* detected using AFLP markers. *Journal of Evolutionary Biology*, 14, 611–619. <https://doi.org/10.1046/j.1420-9101.2001.00304.x>
- Williams, G. A. (1990). The comparative ecology of the flat periwinkles, *Littorina obtusata* (L.) and *L. mariae* Sacchi et Rastelli. *Field Studies*, 7, 469–482.
- Wood, H. M., Grahame, J. W., Humphray, S., Rogers, J., & Butlin, R. K. (2008). Sequence differentiation in regions identified by a genome

scan for local adaptation. *Molecular Ecology*, 17, 3123–3135. <https://doi.org/10.1111/j.1365-294X.2008.03755.x>

Zhivotovsky, L. A. (1999). Estimating population structure in diploids with multilocus dominant DNA markers. *Molecular Ecology*, 8, 907–913. <https://doi.org/10.1046/j.1365-294x.1999.00620.x>

How to cite this article: Galindo J, Carvalho J, Sotelo G, et al. Genetic and morphological divergence between *Littorina fabalis* ecotypes in Northern Europe. *J. Evol. Biol.* 2021;34:97–113. <https://doi.org/10.1111/jeb.13705>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.